

Component-resolved diagnostics in pet allergy: Current perspectives and future directions



Ann-Marie M. Schoos, MD, PhD,^{a,b} Bright I. Nwaru, PhD,^{c,d} and Magnus P. Borres, MD, PhD^{e,f}
Denmark; and Gothenburg and Uppsala, Sweden

Copenhagen and Slagelse,

Furry mammals kept as pets are important allergen sources. The prevalence of sensitization to dander from various animals appears to be increasing worldwide. Several mammalian allergens from diverse species and distinct protein families have been characterized, and some are available for component-resolved diagnostics (CRD). This review presents an overview of mammalian aeroallergens, with a focus on cat, dog, and horse allergens. The potential of CRD in fine-tuning the diagnostic workup following traditional methods based on whole-allergen extracts and allergen immunotherapy is discussed. The review highlights the clinical utility of CRD, particularly as a marker/predictor of increased asthma risk and disease severity. Finally, several perspectives of the future implications of CRD are offered in the context of furry animal allergens. (*J Allergy Clin Immunol* 2021;147:1164-73.)

Key words: Pet allergy, dog, cat, horse, component-resolved diagnostics, asthma risk

Allergy to cats and dogs has been recognized for many years as a major risk factor for the development of asthma and allergic rhinitis.¹ In Europe and the United States, 24% to 38% of households own dogs and 25% own cats.^{2,3} In the United States, only 0.7% own horses.³ These estimates indicate that there is a high exposure in households to furred animals, in particular cats and dogs, at least in the Western world. Allergens from pet animals are mainly present in their fur, saliva, and urine and are spread into the environment through shedding of hair and dander.⁴ A national survey of 831 random homes in the United States

Abbreviations used

AIT: Allergen immunotherapy
CRD: Component-resolved diagnostics
FENO: Fractional exhaled nitric oxide
sIgE: Specific IgE
SPT: Skin prick test

found that all homes contained dog allergen (100%) and most contained cat allergen (99.9%), even homes without pets.⁵

Over the past decades, the prevalence of allergy to furry animals⁶ and the prevalence of asthma and allergic rhinitis^{7,8} have been increasing. Allergy to cat is the most common mammalian-origin allergy in humans,⁹ and sensitization to cat is found in up to 1 in 5 adults worldwide.¹⁰ The diagnosis of cat allergy has proven rather uncomplicated, probably because most patients react to 1 main protein, Fel d 1.¹¹ The cat skin prick test (SPT) extracts contain mainly Fel d 1, but might differ in content of Fel d 4, which needs further investigation.

The diagnosis of dog allergy is more challenging; self-reporting misclassifies allergic status in many patients,¹² and the protein content in SPT extracts vary up to 20-fold within different manufacturers.¹³

Component-resolved diagnostics (CRD) is beginning to gain greater recognition for pet allergy diagnostics. CRD identifies specific IgE (sIgE) responses to specific molecular targets, and several molecular allergy components have been identified for cat, dog, and horse. However, this dimension also increases the complexity of diagnostics because some components may represent cross-reactive sensitizations.^{14,15}

The aim of this review was to provide an overview of the symptomatology and epidemiology of pet allergy. The focus is on CRD for cat, dog, and horse allergens, with a description of the different proteins, frequency of sensitization, and the clinical utility of pet components. A search was performed in PubMed to identify studies on CRD for pet components (cat, dog, and horse allergen components) published between 1997 and mid-2020.

PET ALLERGY: FREQUENCY, TREND, AND IMPACT ON ALLERGIC DISEASES

An international survey of more than 27,000 participants from 22 countries estimated that 57% of the population has at least 1 pet at home, most commonly dogs (33%) and cats (23%)¹⁶ (Fig 1). A survey of almost 13,000 German children reported a sensitization rate of 12.6% to animal dander. The prevalence increased with age from 5.7% in 3- to 6-year-olds to 11.5% in 7- to 10-year-olds, and reached 17.2% in 14- to 17-year-olds.¹⁷ A Swedish birth cohort study of more than 4000 children reported a similar increase in sensitization rates to horse, cat, and dog from

From ^aCopenhagen Prospective Studies on Asthma in Childhood (COPSAC), Herlev and Gentofte Hospital, University of Copenhagen, Copenhagen; ^bthe Department of Pediatrics, Slagelse Sygehus, Slagelse; ^cthe Krefting Research Centre, Institute of Medicine, and ^dthe Wallenberg Centre for Molecular and Translational Medicine, University of Gothenburg; ^eThermo Fisher Scientific, Uppsala; and ^fthe Department of Maternal and Child Health, Uppsala University, Uppsala.

B.I.N. is supported by the Knut and Alice Wallenberg Foundation, the Wallenberg Centre for Molecular and Translational Medicine, and the VBG Group Herman Krefting Foundation on Asthma and Allergy. The funding agencies of B.I.N. did not have any role in design and conduct of the study; collection, management, and interpretation of the data; or preparation, review, or approval of the manuscript.

Disclosure of potential conflict of interest: M. P. Borres is an employee of Thermo Fisher Scientific. The rest of the authors declare no potential, perceived, or real conflict of interest regarding the content of this manuscript.

Received for publication October 7, 2020; revised November 30, 2020; accepted for publication December 23, 2020.

Available online January 11, 2021.

Corresponding author: Ann-Marie M. Schoos, MD, PhD, COPSAC, Copenhagen Prospective Studies on Asthma in Childhood, Gentofte and Herlev Hospital, University of Copenhagen, Ledreborg Alle 34, 2820 Gentofte, Denmark. E-mail: ann-marie.schoos@dbac.dk.

The CrossMark symbol notifies online readers when updates have been made to the article such as errata or minor corrections

0091-6749/\$36.00

© 2021 American Academy of Allergy, Asthma & Immunology
<https://doi.org/10.1016/j.jaci.2020.12.640>

4 to 20 years, reaching 9.8%, 19.6%, and 17.0%,^{18,19} respectively. High prevalence of sensitization to common airborne allergens, including cats and dogs, has also been observed in adults,²⁰ although the prevalence in adults is lower and monosensitization is more frequent than in children.²¹ In Brazil, sensitization to furry animals, especially to dog, increased dramatically among allergic children and adolescents between 2004 and 2016, seemingly reflecting a sedentary “indoor lifestyle.”²²

Geographic variation in the prevalence of allergic sensitization to furry animals has been attributed to cultural differences, environmental factors, and rate of pet ownership.^{23,24} A large international multicenter study in adults reported a sensitization rate to cat of 8.8% (range, 1.2%–22.4%) as measured by SPT¹⁰ and the sensitization rate for dog was 20.4% among adult Korean subjects.²⁵ Approximately 26% of European adults coming to the clinic for suspected allergy to inhalant allergens are sensitized to cat and 27% to dog, according to another large patient-based study of SPT for aeroallergens (Global Asthma and Allergy European Network).^{24,26}

The severity of induced symptoms of pet allergy varies widely from discomfort associated with rhinitis and conjunctivitis to severe asthma, which can develop into a life-threatening condition.¹ Some studies have found an association between sensitization to cat and dog in early childhood and later development of asthma and allergic rhinitis.^{27,28} Another study found no association between early cat and dog exposure and later development of allergic rhinitis to cat and dog, respectively,²⁹ and one even found neonatal exposure to dog to reduce the risk of atopic dermatitis in early childhood.³⁰ A number of pet animal components, mostly produced as recombinant proteins, are now available for CRD, offering improved diagnostic workup (Fig 2), particularly in patients with polysensitization and/or severe asthma.^{4,31} There is little evidence of the actual value of sIgE and SPTs in dog and cat allergy, and large variability between content of skin-prick extracts has been documented.^{13,32} However, the ability of pet allergen CRD can help distinguish between sensitizations specific to singular species and sensitizations due to cross-reactivity.³¹

ALLERGENS AND SENSITIZATION PATTERNS

The World Health Organization/International Union of Immunological Societies³³ lists 36 major mammalian-derived aeroallergens (Table I). They can be classified into the following protein families: lipocalins, secretoglobins, serum albumins, kallikreins, and latherins.^{19,23} A major allergen is defined as one to which more than 50% of patients with an allergy to its source react.¹⁸

Lipocalins

The most important group of mammalian aeroallergens is lipocalins.³⁴ These allergens are produced in secretory glands and are present in skin, urine, saliva, sweat, and sebum.³⁴ At least 1 allergen from the lipocalin protein family has been identified in each species (Table I). From the lipocalin family, cats contain Fel d 4 and Fel d 7. Studies have shown that saliva is the main source of these allergens, and that they are deposited through grooming on the fur.^{35,36} Fel d 4 levels showed no relation to hair length; however, neutered female cats had significantly higher levels compared with not neutered cats (17.4 vs 2.2 µg/g;

$P = .039$). IgE reactivity to cat allergen Fel d 4 has been observed in up to 63% of subjects allergic to cat,³⁷ and IgE reactivity to Fel d 7 has been reported in 38%.³⁶ Fel d 7 has high potential to cross-react with Can f 1, with which it shares 62% amino acid identity.¹⁰

Dogs contain Can f 1, Can f 2, Can f 4, and Can f 6 from the lipocalin family. Dog studies have shown that higher lipocalin levels were detected in saliva than in fur, with Can f 4 showing the highest concentration.³² Sensitization to Can f 4 has been found in up to 46% of patients allergic to dog.³⁸ Can f 2 was exclusively found in saliva and is characterized as a minor allergen, with sensitization present in about 20% to 33% of patients allergic to dog.^{1,39,40} Among patients allergic to dog, 50% to 90% have antibodies to Can f 1, 20% to 33% to Can f 2,^{1,39,40} 35% to 46% to Can f 4,^{38,41} and 56% to Can f 6.⁴²

Two horse lipocalins, Equ c 1 and Equ c 2, have been identified. In an adult Swedish population, 2% were sensitized to Equ c 1 and 12% among patients with asthma.²¹ Up to 76% of patients with horse allergy are sensitized to Equ c 1.⁴³

The lipocalins comprise a diverse protein family with specific patterns of cross-reactivity among certain of its members, for example, between Can f 6 and Fel d 4 and Equ c 1,^{44–46} Mus m 1 and Rat n 1,⁴⁷ Equ c 1 and Mus m 1,⁴³ and Fel d 7 and Can f 1.⁴⁸ Lipocalin allergens Equ c 1, Fel d 4, Can f 6, and Mus m 1 show sequence identities between 47% and 69% (Fig 3, A).¹⁸

Secretoglobins

Two mammalian allergens have been categorized as members of the secretoglobin protein family, namely Fel d 1 from cat and Ory c 3 from rabbit.⁴³ Fel d 1 is mainly produced in sebaceous and salivary glands and is transferred to fur by grooming.⁴⁸ Fel d 1 is the most important allergen in cat allergy, shown to react with IgE from 90% of cat-sensitized individuals, and to account for up to 90% of IgE reactivity to cat dander.⁴⁹ Sequence identity between Fel d 1 and Ory c 3 is very low, and no IgE cross-reactivity has been observed.⁴⁷

Serum albumins

According to the World Health Organization/International Union of Immunological Societies allergen nomenclature database,³³ 6 mammalian serum albumin allergens have been identified, including Bos d 6 (domestic cattle), Can f 3 (dog), Cav p 4 (guinea pig), Equ c 3 (domestic horse), Fel d 2 (cat), and Sus s 1 (domestic pig). Serum albumins are highly abundant proteins present in blood, dander, milk, and other secretions, but are considered minor allergens with low prevalence of IgE reactivity among patients allergic to its source. Serum albumins remain relevant because they are responsible for species cross-reactivity due to high sequence identity (up to 82%),⁵⁰ for example, between cat and pig (Fig 3, B). In a group of 39 highly sensitized patients allergic to cat, 23% had sIgE to Fel d 2 and more than half of these had sIgE to Sus s 1.⁵¹ Monosensitization to serum albumins seems rare. Sensitization to serum albumins is in most cases observed in combination with sIgE directed against major allergens.⁴⁴ Serum albumins may play a significant role as cross-reacting allergens in individuals sensitized to dander of multiple animal species. These mechanisms could explain why some atopic patients develop allergic sensitization to mammalian allergens in the absence of contact with the animals.⁵² Indeed, because of cross-reactivity, children with persistent milk allergy

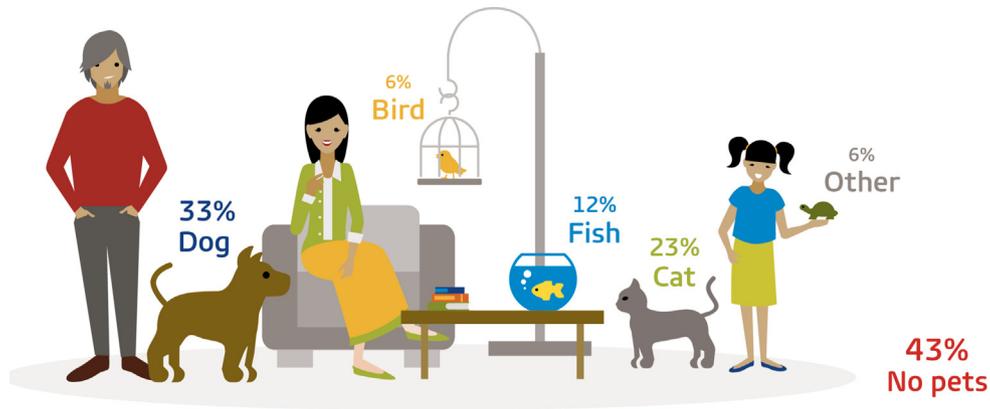


FIG 1. Percentage of people living with pets (Growth from Knowledge survey of >27,000 internet users in 22 countries).¹⁶

and BSA sensitization show an increased risk of allergy to animal dander with symptoms of rhinoconjunctivitis and asthma.⁵³

Kallikrein

Can f 5 is so far the only identified allergen from the kallikrein protein family. The protein is secreted from the prostate and therefore present only in male dogs. The allergen can be isolated from the urine of male dogs, but it is also present in dog hair and dander.⁵⁴ Among patients allergic to dogs, 31% to 70% showed IgE reactivity to Can f 5, and up to 58% of these patients were monosensitized to Can f 5 alone⁵⁴⁻⁵⁹ (Table II). Can f 5 has also been recently reported as the most common dog component sensitization in a Swedish adult population.²¹ The Can f 5 amino acid sequence shows no significant similarity to any known animal dander or urinary allergen.⁴ Therefore, monosensitization to Can f 5 could be a highly specific marker for allergy to male dogs. Schoos et al^{64,65} have recently shown that patients allergic to dogs who are truly monosensitized to Can f 5 are allergic only to male dogs. These patients showed no reaction to SPT or conjunctival allergen provocation test using female dog extract and thereby tolerated female dogs. On the contrary, Käck et al⁶¹ found no correlation between nasal provocation test with dog dander extract and monosensitization to Can f 5. This can be due to the dog extract containing a mix of all the dog components (from both male and female dogs), and possibly very low levels of Can f 5. Can f 5 is a 28-kDa protein prostatic kallikrein that cross-reacts with prostate-specific antigen of human seminal plasma.⁵⁴ Because these 2 proteins share substantial structural similarity, 60% sequence identity, it is possible that sensitization to Can f 5 in women might lead to allergic reactions to human seminal fluid during intercourse.⁶⁶⁻⁶⁹

Latherins

Two allergens belonging to the latherin protein family, namely Equ c 4 and Fel d 8, have been identified. Equ c 4 is an abundant protein constituent in sweat, saliva, and dander of horses.^{70,71} An IgE-binding frequency to Equ c 4 of 77% in horse-sensitized subjects has been reported.⁷² Fel d 8 has been found in saliva of cats; however, with an IgE-binding frequency of only 19% among individuals allergic to cat, it is not considered a major cat allergen.³⁶

CLINICAL UTILITY OF CRD

Allergy diagnosis is supported by the detection of allergen sIgE antibodies using whole extracts or individual allergenic proteins from animal dander. However, component-based IgE testing can distinguish a primary sensitization from a cross-sensitization to a higher extent than whole-allergen extracts.¹⁷ Although the possibility of using single components to replace whole extracts in daily diagnostic practice has been widely debated,^{72,73} a dominating view is that extracts are still needed as first-line testing to ensure the detection of patients sensitized to components other than those that are available for CRD.

The availability of allergen component tests has driven new epidemiological studies to analyze the prevalence and clinical relevance of sIgE directed against individual molecular allergens. Thus, CRD may be useful in detecting atypical sensitization profiles that involve minor allergen components. It can also be used to describe patient-specific IgE profiles to establish predictive risk markers and to develop strategies for therapeutic intervention.^{1,44} When clinical history and investigations are inconclusive, molecular allergology can add valuable clinical information in the diagnostic workup (Fig 2).¹² CRD may further prove useful in terms of improved clinical sensitivity in cases in which a relevant allergen is scarcely represented in the natural allergen extract, as is the case of PR-10 allergens in certain plant foods such as fruits and nuts. Superior clinical specificity of component IgE testing compared with whole-extract testing has been demonstrated for allergies to several foods, including peanut,⁷⁴ hazelnut,^{75,76} and cashew nut.^{77,78}

PET ALLERGEN COMPONENTS AS MARKERS OF INCREASED DISEASE RISK AND DISEASE SEVERITY

CRD could provide markers of increased asthma risk. Sensitization to Equ c 1 has been associated with severe childhood asthma.³¹ Furthermore, asthmatic children with cat allergy have higher Fel d 1 sIgE levels than children with rhinitis only.⁷⁹ Another study of 696 Swedish children allergic to cat found that asthma symptoms on contact with cat were significantly associated with sIgE to cat allergens Fel d 1 and Fel d 4.⁵⁷ Among dog-sensitized children, most were sensitized to more than 1 dog component, and cosensitization to Can f 5 and Can f 1/2 conferred

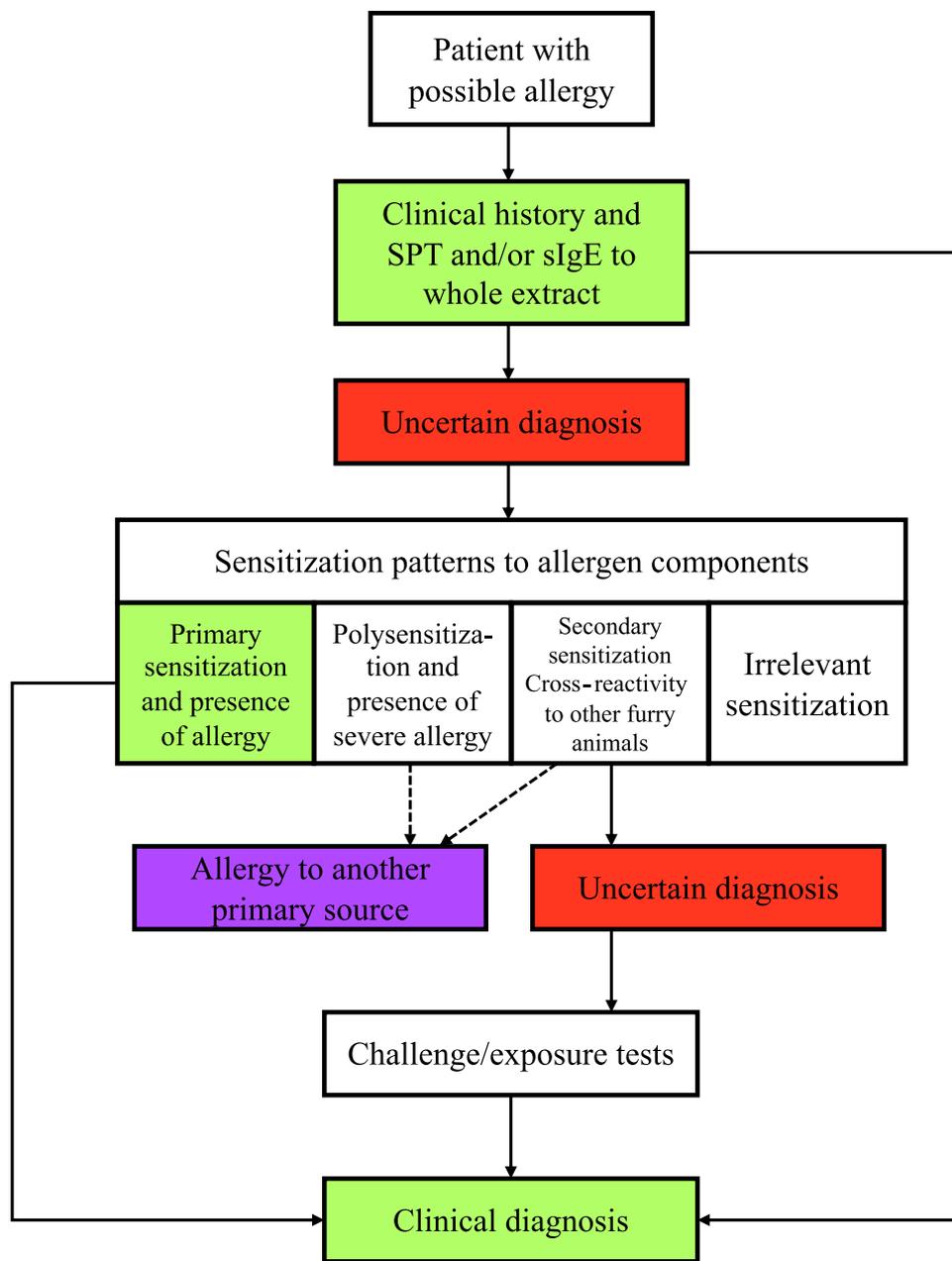


FIG 2. Algorithm for diagnostic workup of patients allergic to animal dander (adapted from Gerth van Wijk¹²).

the greatest risk for asthma.⁵⁷ Furthermore, the study confirmed that asthma was associated with higher levels of component sensitization. Progression of allergic sensitization over time has been shown to involve IgE recognition of an increasing number of components from the sensitizing allergen source, forming the basis for the concept of molecular spreading, in which sensitization to a greater number of components from the same allergen source correlates with disease severity.⁸⁰ A recent study reported that asthmatic pediatric patients with IgE to Fel d 2 serum albumin and Fel d 4 and Fel d 7 lipocalins were more likely to have persistent type 2 inflammation.⁸¹ A longitudinal study of 398 children found that sensitization to dog, cat, and horse throughout

childhood was significantly associated with asthma at age 7 years.⁸² Similarly, a small early-stage study comparing children with severe asthma *versus* children with controlled asthma demonstrated that those with severe asthma had higher levels of IgE antibodies toward cat, dog, and horse components.³¹ Along those lines, a cross-sectional cohort study of 269 children found that sensitization to members of the lipocalin protein family, mainly derived from cat, dog, and horse, was associated with current asthma.⁸³ The relationship between sensitization to specific allergen components and disease has been investigated by Simpson et al,⁸⁴ who identified patterns of response to allergen component groups and investigated associations with asthma in children.

TABLE I. Pet allergen components (www.allergen.org)

Animal	Component	Protein type
Domestic cattle	Bos d 2	Lipocalin
	Bos d 3	S100 calcium-binding protein A7
Dog	Can f 1*	Lipocalin
	Can f 2*	Lipocalin
	Can f 3*	Serum albumin
	Can f 4*	Lipocalin
	Can f 5*	Arginine esterase, prostatic kallikrein
	Can f 6*	Lipocalin
Guinea pig	Can f 7	Epididymal secretory protein E1, or Niemann Pick type C2 protein
	Cav p 1	Lipocalin
	Cav p 2	Lipocalin
	Cav p 3	Lipocalin
	Cav p 4	Serum albumin
Donkey	Cav p 6	Lipocalin
	Equ a 6	Lysozyme
Domestic horse	Equ c 1*	Lipocalin
	Equ c 2	Lipocalin
	Equ c 3*	Serum albumin
	Equ c 4	Latherin
Cat	Equ c 6	Lysozyme
	Fel d 1*	Secretoglobin (uteroglobin, chain 1)
	Fel d 2*	Serum albumin
	Fel d 3	Cystatin
	Fel d 4*	Lipocalin
	Fel d 5w	IgA
	Fel d 6w	IgM
	Fel d 7*	Lipocalin (Von Ebner gland protein)
Golden hamster Syrian hamster	Fel d 8	Latherin-like protein
	Mes a 1	Lipocalin
Mouse	Mus m 1*	Lipocalin/urinary prealbumin
Rabbit	Ory c 1	Lipocalin
	Ory c 3	Secretoglobin (lipophilin)
	Ory c 4	Lipocalin
Siberian hamster	Phod s 1	Lipocalin
Rat	Rat n 1	Alpha-2u-globulin/lipocalin
Domestic pig	Sus s 1	Serum albumin

*Mammalian allergens currently available for CRD.

Sensitization to a group that included 27 components of plant, animal, and fungal origin from 12 protein families was most strongly associated with asthma and decreased lung function (lower FEV₁, $P < .001$).⁸⁴ Similar results have been reported in several subsequent studies.^{60,85} In another study, polysensitization to 3 or more animal-derived components (lipocalins, kallikrein, and secretoglobin) was associated with severe asthma, increased bronchial inflammation, and a trend toward more courses of oral corticosteroid treatment.⁸⁶ Similar findings were recently reported in a study of dog-sensitized children,⁶¹ where Käck et al⁶¹ found an association between sensitization to an increasing number of dog allergen components and a positive nasal challenge result.

Other than asthma, high levels of IgE antibodies to Fel d 4 and also Fel d 2 have been associated with atopic dermatitis in children with cat allergy.⁸⁷ This is the only study found so far regarding measurements of IgE antibodies to pet components in individuals with atopic dermatitis. It is an interesting finding that needs to be verified by larger studies.

The use of allergen components compared with whole extracts as predictors of disease severity was evaluated by Asarnoj et al⁶⁰ in a large longitudinal population-based pediatric study.

Sensitization to Fel d 1 and Can f 1 at age 4 years and molecular polysensitization to cat or dog components predicted allergy to cat and dog at age 16 years significantly better than did IgE to whole cat or dog extracts. In addition, Patelis et al⁷² showed that adults sensitized to both cat extract and 1 or more of cat components Fel d 1, Fel d 2, and Fel d 4 had higher fractional exhaled nitric oxide (FENO) ($P = .008$) and more bronchial responsiveness ($P = .002$) than subjects sensitized to the extract but not to any of the cat components tested. Furthermore, subjects who were sensitized to the tested cat components were more likely to develop asthma ($P = .005$) and rhinitis ($P = .007$) over a 12-year period than those who were not, highlighting the value of CRD in predicting disease severity.⁷² Another study of adults found that sensitization to furry animal allergen components, sensitization patterns, and clusters were associated with a substantially increased risk of asthma, rhinitis, and concomitant asthma/rhinitis. Sensitization to Fel d 1, Can f 1, Can f 2, and Can f 3 and polysensitization (ie, sensitization to more than 2 components) were further associated with increased FENO and eosinophil levels. The impact of polysensitization was also confirmed when a data-driven approach was used to cluster sensitization to the allergen components. Here, individuals who fell into the

A Lipocalins

	Bos d 23k	Bos d 2	Mus m 1	Equ c 2	Equ c 1	Fel d 7	Fel d 4	Can f 6	Can f 4	Can f 2	Can f 1
Can f 1	22	26	21	23	28	63	26	26	24	24	100
Can f 2	24	20	26	26	26	23	25	24	26	100	
Can f 4	37	32	28	35	29	23	27	26	100		
Can f 6	28	27	47	30	57	24	69	100			
Fel d 4	28	31	50	31	68	20	100				
Fel d 7	21	23	21	23	26	100					
Equ c 1	28	33	47	34	100						
Equ c 2	46	32	28	100							
Mus m 1	27	30	100								
Bos d 2	33	100									
Bos d 23k	100										

B Serum albumins

	Sus s 1	HSA	Fel d 2	Equ c 3	Cav p 4	Can f 3	Bos d 6
Bos d 6	79	76	78	74	70	76	100
Can f 3	78	80	87	76	73	100	
Cav p 4	72	72	76	72	100		
Equ c 3	76	76	78	100			
Fel d 2	79	82	100				
HSA (human serum albumin)	75	100					
Sus s 1	100						

FIG 3. Amino acid sequence identity (adapted from Matricardi et al¹⁸) for (A) lipocalins and (B) serum albumins.

“multiple sensitization” cluster were at increased risk of asthma and rhinitis and had higher levels of FENO and eosinophils than those in other clusters. Thus, sensitization to furry animal allergen components is an important predictor of asthma outcome and an indicator of severity.⁸⁸ Taken together, available data indicate that measuring component sIgE offers insights into the progression and severity of asthma and allergic rhinitis.

New clinical implications and future perspectives

Monosensitization to Can f 5 could be a highly specific marker for allergy to male dogs. The only study supporting this theory so far is the Danish study by Schoos et al that shows that individuals monosensitized to Can f 5 show different reactions to male and female dog allergen provocation.^{64,65} Their findings are indirectly supported by Liccardi et al⁶² who found that monosensitization to Can f 5, as well as high levels of IgE to Can f 5, was associated with a prevalent exposure to male dog. The authors conclude that for such patients, the correct prescription of allergen immunotherapy (AIT) and the possibility to own a female dog should be considered. In theory, neutering the male dog could also be a possibility to stop the production of Can f 5; however, this has never been tested in practice. The low cross-reactivity between Can f 5 and the other

dog components⁴ explains why patients monosensitized to Can f 5 did not react on exposure to female dog extracts.^{64,65} Moreover, the possibility of developing an allergy to female dogs is not likely to be increased. An epidemiological study from Sweden studied whether different dog characteristics modify the risk of asthma among children (n = 23,585) exposed to dogs during their first year of life.⁸⁹ Children exposed to female dogs had a lower risk of asthma compared with those exposed to male dogs. They were also able to analyze the impact of the breed and interestingly they found that exposure to so-called hypoallergenic dogs was associated with a higher risk of allergy; however, this association became nonsignificant when adjusting for parental allergy, but still with a trend of an association (odds ratio, 1.20; 95% CI, 0.98-1.47). Monosensitization to Can f 5 has been based on evaluation of 4 dog components (Can f 1, 2, 3, and 5), and it varies in frequencies (Table II). IgE to Can f 4 and Can f 6 are now commercially available,⁹⁰ which changes the definition of monosensitization in future studies, and the “true” number of patients monosensitized to Can f 5 will be lower than today’s estimate.

It is perceived that sensitization to Can f 5 confers an increased risk of development of allergic reactions to human seminal fluid, but that proposition needs to be elucidated. A number of case reports have been published supporting this hypothesis.⁵⁴⁻⁵⁶ In

TABLE II. Can f 5 monosensitization in different populations

Year	Country	Characteristics of studied population	Can f 1	Can f 5	Mono Can f 5*	n†	Reference
2009	Spain, the United States, and Sweden	Dog-allergic, adult patients	57%	70%	33%	37	54
2015	Sweden	General cohort, age 11-12 y, n = 219 positive dog sensitization	39%	47%	10%	696	57
2016	Spain	Asthma/rhinitis + positive dog/cat/horse sensitization	66%	33%	19.5%	159	55
2016	Sweden	General cohort, age 16 y	6%	12.6%	56%	779	60
2016	Poland	Allergy patients mean age 35 y + positive pet sensitization	40%	33%	17%	70	56
2017	Spain	Dog-allergic, age 10-61 y	41%	67%	37%	70	58
2017	Japan	Food allergy cohort, age 13 mo, n = 111 positive dog sensitization	81%‡	36%	4%	304	28
2018	Sweden	Patients allergic to dog, age 10-18 y and positive dog sensitization	65%	62%	12%	60	61
2020	Italy	Allergy patients tested with ISAC, age 4-64 y	59%	69%	58%	1403	62
2019	Sweden	Random population, age 16-75 y	2.9%‡	3.3%	49%	1103	21
2019	Sweden	Asthma, age 16-75 y	14%‡	8.7%	37%	769	21
2020	Czech Republic	Allergy patients tested with ISAC, age 1-68 y	13.9%	16.4%	9.8%	1255	63

*Percentage of monosensitization is calculated from the population sensitized to dog extract but not the components Can f 1-3.

†The numbers extracted from the respective population or through communication with authors.

‡Sensitization to Can f 1 or Can f 2.

view of the high number of patients sensitized to Can f 5 and the potential consequences in females, González-de-Olano et al⁶⁹ studied the prevalence of allergic symptoms to human seminal plasma during intercourse among 27 women sensitized to Can f 5 with a male dog at home. They found a high prevalence of allergy to human seminal plasma among this group of women, and it seemed that the presence of allergic symptoms was related to the level of Can f 5. Based on these findings, it would be appropriate to ask women with higher titers of sIgE to Can f 5 about symptomatology during intercourse apart from respiratory symptoms.

In a relatively new approach, the introduction of anti-Fel d 1 immunoglobulin Y in cat food has shown a reduction in immunologically active Fel d 1 in cat saliva, hair, and dander, which in turn leads to reduction in Fel d 1 level in the environment.⁹¹ This provides an opportunity for Fel d 1-sensitized cat owners to treat their cats to reduce their cat-allergic symptoms. Hypothetically, this treatment of the cat would work only if the cat owner is monosensitized to Fel d 1. If the allergic cat owner is polysensitized to Fel d 2, Fel d 4, or Fel d 7, the owner would still be having reactions. The effect of this new interesting treatment needs to be explored further.

Multiplex CRD tests do not only provide detailed information on the patient's sensitization profile in the clinic but are used in clinical and epidemiological research. The number of peer reviewed publications in PubMed has constantly increased over the years. By December 2019, 288 articles were posted on PubMed using the ImmunoCAP ISAC microarray chip technology. CRD has proven to be a useful tool in describing pet sensitization profiles on a population level and thus provides an opportunity to make comparisons across populations. Yamamoto-Hanada et al⁹² found that IgE sensitization to Fel d 1 was found in 26% at age 9 years in an unselected Japanese cohort. This finding is in contrast with an unselected Swedish cohort of adolescents in which 16% of subjects were sensitized to Fel d 1. Correspondingly, 10% versus 4% were sensitized to Can f 1 in the Japanese versus Swedish cohort and 2% versus 3% were sensitized to Equ c

1.⁹³ CRD arrays for multiple components generate complex and rich data sets, which warrants application of novel techniques such as machine learning to analyze the data. These methodologies have identified clusters of component sIgEs to multiple allergenic proteins associated with high risk of asthma in cross-sectional and longitudinal studies. For example, Roberts et al⁹⁴ applied clustering methods to identify co-occurring patterns of components and patterns of sensitization among participants. Furthermore, they used network analysis techniques to explore the connectivity structure of component sIgE, and differences in component sIgE interactions between patients with severe and mild/moderate asthma. They found that participants with severe asthma had higher connectivity among components (ie, more connections between different components), but these connections were weaker. The mild/moderate network had fewer connections, but the connections were stronger. Interestingly, connections among animal components showed higher correlations in severe asthma in both adults and children than in mild to moderate asthma. This analysis provides a strong indication that computational data analysis will be essential to uncovering the nuances of complex sensitization patterns of emerging CRD-based allergens of furry animals, which will constitute important steps for future research.

The chip-based multiplex assay provides interpolate results from an internal calibration curve into semi-quantitative estimates of the IgE antibody as classes or grades.⁹⁵ Their analytical sensitivity is generally less than that of singleplex tests. Because of the low density of allergen on the chip dots, nanogram levels of IgE antibody binding can be interfered with ISAC by the presence of microgram quantities of allergen-specific IgG "blocking" antibodies. The optimal application for these assays resides in epidemiologic studies where they can define the atopic status of different populations in geographically diverse regions.

Treatment options for individuals who are allergic to furry animals include allergen avoidance, medication, and AIT.¹⁸ The ability of components to distinguish primary sensitization from cross-sensitization is important when immunotherapy is

envisaged, to choose the primary sensitizing allergen source for therapy. The availability of CRD has raised the possibility of better-targeted AIT, which up to date is the only treatment able to change the natural course of allergic disease. There is a clinical need for improved extracts for pet allergies because both standardized and unstandardized extracts are used for immunotherapy today in the United States.⁹⁶ As an example, unstandardized dog allergen extracts exist in the United States in 2 formulations—conventional and “acetone-precipitated.” As with all unstandardized allergen extracts, no valid potency measure exists for these dog extracts. In spite of this, some reports and experiences from clinical practice suggest that the use of the acetone-precipitated product will yield better clinical results associated with their higher Can f 1 levels.⁹⁷ These suggestions are not based on clinical studies, and a recent report highlights that the immune response to dog is more complex than previously thought.⁶¹ The medical literature on the use of dog extract immunotherapy in patients with hypersensitivity to dog shows poor and conflicting results of clinical efficacy. This has been attributed to poor-quality extracts and the inherent complex allergenic profile of dogs, which remains without a clearly dominant allergen.⁹⁸ A recent study showed that the molecular profile (number of allergens to which the patients were sensitized or the quantitative value of sIgE to allergens) was not related to response to subcutaneous immunotherapy with cat or dog extracts.⁹⁹ Research into the optimal organism in which to produce recombinant proteins continues. In the future, more potent and patient-specific extracts could be developed from a mixture of organic sources (saliva, urine, and dander) and then hybrid extracts with natural and recombinant allergens could be produced for use in immunotherapy. Component testing and patient-specific hybridized extracts could identify and provide the best therapeutic match to the patient’s pattern of sensitization and therefore may offer better chances for effective therapy.

Based on this review, possible future clinical utility of CRD when a patient presents with suspected dog allergy would be measuring all the dog components (Can f 1, 2, 3, 4, 5, and 6) to clarify monosensitization to Can f 5 (female dog tolerance).⁶⁵ Furthermore, the more components a patient is sensitized to, the higher the likelihood of a clinical reaction when exposed to dog.^{60,61} Finally, higher levels of dog component sensitization and cosensitization to Can f 5 and Can f 1/2 are associated with current asthma.⁵⁷ In patients suspected with cat allergy, Fel d 1, 2, and 4 seem to be the most important components to measure. Sensitization to Fel d 1 is associated with asthma,^{57,79} and polysensitization (Fel d 1, 2, and 4) is associated with both clinical reactivity to cat⁶⁰ and also bronchial responsiveness and increased FENO.⁶⁹ Finally, sensitization to Fel d 2 and Fel d 4 is associated with atopic dermatitis.⁸⁷ In patients with suspected horse allergy, only sensitization to Equ c 1 has been found clinically relevant, because it is associated with asthma.³¹

Conclusions

CRD is allowing detailed analysis of patients’ sensitization profiles and may ultimately facilitate individualized treatments and patient management options. Some questions however remain to be addressed on the clinical utility of CRD in patients with allergy to furry animals.¹ In the interim, CRD should be considered complementary to extract-based testing rather than a replacement (Fig 2). The role of allergenic molecules as markers

and predictors of disease severity needs to be further explored. Further studies are also needed to understand whether CRD could be used to identify patients who are most likely to respond to AIT. CRD may also have a role in monitoring treatment responses following immunotherapy. Debate in the allergy community reveals that the initial skepticism regarding precision allergy molecular diagnostic applications has moved toward the possibility that it could replace SPT in the future.⁹⁰ This concern exists especially in areas debating the preparation of SPT reagents in accordance with Good Manufacturing Procedures.

Overall, CRD has a role in developing patient-tailored treatment that could reduce health care costs, save time for patients, reduce adverse effects, and improve patient quality of life.

REFERENCES

- Konradsen JR, Fujisawa T, van Hage M, Hedlin G, Hilger C, Kleine-Tebbe J, et al. Allergy to furry animals: new insights, diagnostic approaches, and challenges. *J Allergy Clin Immunol* 2015;135:616-25.
- FEDIAF. Statistics. Available at: <http://www.fediaf.org/who-we-are/european-statistics.html>. Accessed September 24, 2020.
- American Veterinary Medical Association. U.S. pet ownership statistics. Available at: <https://www.avma.org/resources-tools/reports-statistics/us-pet-ownership-statistics>. Accessed September 24, 2020.
- Zahradnik E, Raulf M. Respiratory allergens from furred mammals: environmental and occupational exposure. *Vet Sci* 2017;4:1-17.
- Arbes SJ, Cohn RD, Yin M, Muilenberg ML, Friedman W, Zeldin DC. Dog allergen (Can f 1) and cat allergen (Fel d 1) in US homes: results from the National Survey of Lead and Allergens in Housing. *J Allergy Clin Immunol* 2004;114:111-7.
- Rönmark E, Bjerg A, Perzanowski M, Platts-Mills T, Lundbäck B. Major increase in allergic sensitization in schoolchildren from 1996 to 2006 in northern Sweden. *J Allergy Clin Immunol* 2009;124:357-63.e1-15.
- Asher MI, Montefort S, Björkstén B, Lai CKW, Strachan DP, Weiland SK, et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet* 2006;368:733-43.
- Chan SK, Leung DYM. Dog and cat allergies: current state of diagnostic approaches and challenges. *Allergy Asthma Immunol Res* 2018;10:97-105.
- Morris DO. Human allergy to environmental pet danders: a public health perspective. *Vet Dermatol* 2010;21:441-9.
- Bousquet P-J, Chinn S, Janson C, Kogevinas M, Burney P, Jarvis D, et al. Geographical variation in the prevalence of positive skin tests to environmental aeroallergens in the European Community Respiratory Health Survey I. *Allergy* 2007;62:301-9.
- van Ree R, van Leeuwen WA, Bulder I, Bond J, Aalberse RC. Purified natural and recombinant Fel d 1 and cat albumin in vitro diagnostics for cat allergy. *J Allergy Clin Immunol* 1999;104:1223-30.
- Gerth van Wijk R. Diagnosis of dog allergy: beware of the dog. *J Allergy Clin Immunol* 2018;142:1058-9.
- Curin M, Reininger R, Swoboda I, Focke M, Valenta R, Spitzauer S. Skin prick test extracts for dog allergy diagnosis show considerable variations regarding the content of major and minor dog allergens. *Int Arch Allergy Immunol* 2011;154:258-63.
- Valenta R, Lidholm J, Niederberger V, Hayek B, Kraft D, Grönlund H. The recombinant allergen-based concept of component-resolved diagnostics and immunotherapy (CRD and CRIT). *Clin Exp Allergy* 1999;29:896-904.
- Liccardi G, Bilò MB, Manzi F, Piccolo A, Di Maro E, Salzillo A. What could be the role of molecular-based allergy diagnostics in detecting the risk of developing allergic sensitization to furry animals? *Eur Ann Allergy Clin Immunol* 2015;47:163-7.
- insights | GfK Global. Available at: <https://www.gfk.com/insights>. Accessed September 24, 2020.
- Schmitz R, Ellert U, Kalcklösch M, Dahm S, Thamm M. Patterns of sensitization to inhalant and food allergens—findings from the German health interview and examination survey for children and adolescents. *Int Arch Allergy Immunol* 2013;162:263-70.
- Matricardi PM, Kleine-Tebbe J, Hoffmann HJ, Valenta R, Hilger C, Hofmaier S, et al. EAACI Molecular Allergology User’s Guide. *Pediatr Allergy Immunol* 2016;27:1-250.

19. Melén E, Bergström A, Kull I, Almqvist C, Andersson N, Asarnoj A, et al. Male sex is strongly associated with IgE-sensitization to airborne but not food allergens: results up to age 24 years from the BAMSE birth cohort. *Clin Transl Allergy* 2020; 10:15.
20. Warm K, Lindberg A, Lundbäck B, Rönmark E. Increase in sensitization to common airborne allergens among adults—two population-based studies 15 years apart. *Allergy Asthma Clin Immunol* 2013;9:20.
21. Suzuki S, Nwaru BI, Ekerljung L, Sjölander S, Mincheva R, Rönmark EP, et al. Characterization of sensitization to furry animal allergen components in an adult population. *Clin Exp Allergy* 2019;49:495-505.
22. Aranda CS, Cocco RR, Pierotti FF, Mallozi MC, Franco JM, Porto A, et al. Increased sensitization to several allergens over a 12-year period in Brazilian children. *Pediatr Allergy Immunol* 2018;29:321-4.
23. Liccardi G, Triggiani M, Piccolo A, Salzillo A, Parente R, Manzi F, et al. Sensitization to common and uncommon pets or other furry animals: which may be common mechanisms? *Transl Med UniSa* 2016;14:9-14.
24. Heinzerling LM, Burbach GJ, Edenharter G, Bachert C, Bindslev-Jensen C, Bonini S, et al. GA(2)LEN skin test study I: GA(2)LEN harmonization of skin prick testing: novel sensitization patterns for inhalant allergens in Europe. *Allergy* 2009;64:1498-506.
25. Park Y-B, Mo E-K, Lee J-Y, Kim J-H, Kim C-H, Hyun I-G, et al. Association between pet ownership and the sensitization to pet allergens in adults with various allergic diseases. *Allergy Asthma Immunol Res* 2013;5:295-300.
26. Dávila I, Domínguez-Ortega J, Navarro-Pulido A, Alonso A, Antolín-Amerigo D, González-Mancebo E, et al. Consensus document on dog and cat allergy. *Allergy* 2018;73:1206-22.
27. Ingram JM, Sporik R, Rose G, Honsinger R, Chapman MD, Platts-Mills TA. Quantitative assessment of exposure to dog (Can f 1) and cat (Fel d 1) allergens: relation to sensitization and asthma among children living in Los Alamos, New Mexico. *J Allergy Clin Immunol* 1995;96:449-56.
28. Nagao M, Borres MP, Sugimoto M, Petersson CJ, Nakayama S, Kuwabara Y, et al. Sensitization to secretoglobin and lipocalins in a group of young children with risk of developing respiratory allergy. *Clin Mol Allergy* 2017;15:4.
29. Schoos A-MM, Chawes BL, Jelding-Dannemand E, Elfman LB, Bisgaard H. Early indoor aeroallergen exposure is not associated with development of sensitization or allergic rhinitis in high-risk children. *Allergy* 2016;71:684-91.
30. Thorsteinsdottir S, Thyssen JP, Stokholm J, Vissing NH, Waage J, Bisgaard H. Domestic dog exposure at birth reduces the incidence of atopic dermatitis. *Allergy* 2016;71:1736-44.
31. Konradsen JR, Nordlund B, Onell A, Borres MP, Grönlund H, Hedlin G. Severe childhood asthma and allergy to furry animals: refined assessment using molecular-based allergy diagnostics. *Pediatr Allergy Immunol* 2014;25:187-92.
32. Wintersand A, Asplund K, Binnmyr J, Holmgren E, Nilsson OB, Gafvelin G, et al. Allergens in dog extracts: implication for diagnosis and treatment. *Allergy* 2019; 74:1472-9.
33. WHO/IUIS Allergen Nomenclature Home Page. Available at: <http://www.allergen.org/>. Accessed September 24, 2020.
34. Hilger C, Kuehn A, Hentges F. Animal lipocalin allergens. *Curr Allergy Asthma Rep* 2012;12:438-47.
35. Kelly SM, Karsh J, Marcelo J, Boeckh D, Stepner N, Santone B, et al. Fel d 1 and Fel d 4 levels in cat fur, saliva, and urine. *J Allergy Clin Immunol* 2018;142:1990-2.e3.
36. Smith W, O'Neil SE, Hales BJ, Chai TLY, Hazell LA, Tanyaratrisakul S, et al. Two newly identified cat allergens: the von Ebner gland protein Fel d 7 and the latherin-like protein Fel d 8. *Int Arch Allergy Immunol* 2011;156:159-70.
37. Smith W, Butler AJL, Hazell LA, Chapman MD, Pomés A, Nickels DG, et al. Fel d 4, a cat lipocalin allergen. *Clin Exp Allergy* 2004;34:1732-8.
38. Rytönen-Nissinen M, Saarelainen S, Randell J, Häyrynen J, Kalkkinen N, Virtanen T. IgE reactivity of the dog lipocalin allergen Can f 4 and the development of a sandwich ELISA for its quantification. *Allergy Asthma Immunol Res* 2015; 7:384.
39. Konieczny A, Morgenstern JP, Bizinkauskas CB, Lilley CH, Brauer AW, Bond JF, et al. The major dog allergens, Can f 1 and Can f 2, are salivary lipocalin proteins: cloning and immunological characterization of the recombinant forms. *Immunology* 1997;92:577-86.
40. Saarelainen S, Taivainen A, Rytönen-Nissinen M, Auriola S, Immonen A, Mäntyjärvi R, et al. Assessment of recombinant dog allergens Can f 1 and Can f 2 for the diagnosis of dog allergy. *Clin Exp Allergy* 2004;34:1576-82.
41. Mattsson L, Lundgren T, Olsson P, Sundberg M, Lidholm J. Molecular and immunological characterization of Can f 4: a dog dander allergen cross-reactive with a 23 kDa odorant-binding protein in cow dander. *Clin Exp Allergy* 2010; 40:1276-87.
42. Wang Y-J, Li L, Song W-J, Zhou Y-J, Cao M-D, Zuo X-R, et al. *Canis familiaris* allergen Can f 6: expression, purification and analysis of B-cell epitopes in Chinese dog allergic children. *Oncotarget* 2017;8:90796-807.
43. Saarelainen S, Rytönen-Nissinen M, Rouvinen J, Taivainen A, Auriola S, Kauppinen A, et al. Animal-derived lipocalin allergens exhibit immunoglobulin E cross-reactivity. *Clin Exp Allergy* 2008;38:374-81.
44. Hilger C, van Hage M, Kuehn A. Diagnosis of allergy to mammals and dish: cross-reactive vs. specific markers. *Curr Allergy Asthma Rep* 2017;17:64.
45. Hilger C, Swiontek K, Arumugam K, Lehnert C, Hentges F. Identification of a new major dog allergen highly cross-reactive with Fel d 4 in a population of cat- and dog-sensitized patients. *J Allergy Clin Immunol* 2012;129:1149-51.
46. Nilsson OB, Binnmyr J, Zoltowska A, Saarne T, van Hage M, Grönlund H. Characterization of the dog lipocalin allergen Can f 6: the role in cross-reactivity with cat and horse. *Allergy* 2012;67:751-7.
47. Jeal H, Harris J, Draper A, Taylor AN, Cullinan P, Jones M. Dual sensitization to rat and mouse urinary allergens reflects cross-reactive molecules rather than atopy. *Allergy* 2009;64:855-61.
48. Apostolovic D, Sánchez-Vidaurre S, Waden K, Curin M, Grundström J, Gafvelin G, et al. The cat lipocalin Fel d 7 and its cross-reactivity with the dog lipocalin Can f 1. *Allergy* 2016;71:1490-5.
49. Bonnet B, Messaoudi K, Jacomet F, Michaud E, Fauquet JL, Caillaud D, et al. An update on molecular cat allergens: Fel d 1 and what else? Chapter 1: Fel d 1, the major cat allergen. *Allergy Asthma Clin Immunol* 2018;14:14.
50. Chruszcz M, Mikolajczak K, Mank N, Majorek KA, Porebski PJ, Minor W. Serum albumins—unusual allergens. *Biochim Biophys Acta* 2013;1830:5375-81.
51. Hilger C, Kohlen M, Grigioni F, Lehnert C, Hentges F. Allergic cross-reactions between cat and pig serum albumin. Study at the protein and DNA levels. *Allergy* 1997;52:179-87.
52. Liccardi G, Asero R, D'Amato M, D'Amato G. Role of sensitization to mammalian serum albumin in allergic disease. *Curr Allergy Asthma Rep* 2011;11:421-6.
53. Vicente-Serrano J, Caballero ML, Rodríguez-Pérez R, Carretero P, Pérez R, Blanco JG, et al. Sensitization to serum albumins in children allergic to cow's milk and epithelia. *Pediatr Allergy Immunol* 2007;18:503-7.
54. Mattsson L, Lundgren T, Everberg H, Larsson H, Lidholm J. Prostatic kallikrein: a new major dog allergen. *J Allergy Clin Immunol* 2009;123:362-8.
55. Uriarte SA, Sastre J. Clinical relevance of molecular diagnosis in pet allergy. *Allergy* 2016;71:1066-8.
56. Ukleja-Sokolowska N, Gawrońska-Ukleja E, Żbikowska-Gotz M, Socha E, Lis K, Sokolowski Ł, et al. Analysis of feline and canine allergen components in patients sensitized to pets. *Allergy Asthma Clin Immunol* 2016;12:61.
57. Bjerg A, Winberg A, Berthold M, Mattsson L, Borres MP, Rönmark E. A population-based study of animal component sensitization, asthma, and rhinitis in schoolchildren. *Pediatr Allergy Immunol* 2015;26:557-63.
58. Basagaña M, Luengo O, Labrador M, Garriga T, Mattsson L, Lidholm J, et al. Component-resolved diagnosis of dog allergy. *J Investig Allergol Clin Immunol* 2017;27:185-7.
59. Villalta D, Milanese M, Da Re M, Sabatino G, Sforza M, Calzetta L, et al. Frequency of allergic sensitization to Can f 5 in North East Italy. An analysis of 1403 ISACs 112 (Component Resolved Diagnosis) collected retrospectively. *Eur Ann Allergy Clin Immunol* 2019;51:186-9.
60. Asarnoj A, Hamsten C, Wadén K, Lupinek C, Andersson N, Kull I, et al. Sensitization to cat and dog allergen molecules in childhood and prediction of symptoms of cat and dog allergy in adolescence: a BAMSE/MeDALL study. *J Allergy Clin Immunol* 2016;137:813-21.e7.
61. Käck U, Asarnoj A, Grönlund H, Borres MP, van Hage M, Lilja G, et al. Molecular diagnostics refine characterization of children sensitized to dog dander. *J Allergy Clin Immunol* 2018;142:1113-20.e9.
62. Liccardi G, Calzetta L, Bilò MB, Brusca I, Cecchi L, Costantino MT, et al. A prevalent exposure to male dog is a risk factor for exclusive allergic sensitization to Can f 5: an Italian multicenter study. *J Allergy Clin Immunol Pract* 2020;8:2399-401.
63. Vachová M, Panzner P, Vlas T, Vítovcová P. Analysis of sensitization profiles in central European allergy patients focused on animal allergen molecules. *Int Arch Allergy Immunol* 2020;181:278-84.
64. Schoos A-MM, Bønnelykke K, Chawes BL, Stokholm J, Bisgaard H, Kristensen B. Precision allergy: separate allergies to male and female dogs. *J Allergy Clin Immunol Pract* 2017;5:1754-6.
65. Schoos A-MM, Chawes BL, Bloch J, Hansen B, Stokholm J, Bønnelykke K, et al. Children monosensitized to Can f 5 show different reactions to male and female dog allergen extract provocation: a randomized controlled trial. *J Allergy Clin Immunol Pract* 2020;8:1592-7.e2.
66. Tanaka M, Nakagawa Y, Kotobuki Y, Katayama I. A case of human seminal plasma allergy sensitized with dog prostatic kallikrein, Can f 5. *Allergol Int* 2019;68: 259-60.

67. Basagaña M, Bartolome B, Pastor-Vargas C, Mattsson L, Lidholm J, Labrador-Horrillo M. Involvement of Can f 5 in a case of human seminal plasma allergy. *Int Arch Allergy Immunol* 2012;159:143-6.
68. Köfler L, Köfler H, Mattsson L, Lidholm J. A case of dog-related human seminal plasma allergy. *Eur Ann Allergy Clin Immunol* 2012;44:89-92.
69. González-de-Olano D, Gandolfo-Cano M, de-Calzada-Bustingorri MP, González-Mancebo E, de-Andrés-Martín A, Cuesta-Herranz J, et al. Prevalence of allergy to human seminal fluid among women with allergy to male dog and sensitization to Can f 5. *Clin Exp Allergy* 2018;48:1368-70.
70. McDonald RE, Fleming RI, Beeley JG, Bovell DL, Lu JR, Zhao X, et al. Latherin: a surfactant protein of horse sweat and saliva. *PLoS One* 2009;4:e5726.
71. Victor S, Binnmyr J, Lampa E, Rask-Andersen A, Elfman L. Levels of horse allergen Equ c 4 in dander and saliva from ten horse breeds. *Clin Exp Allergy* 2019;49:701-11.
72. Patelis A, Gunnbjornsdottir M, Alving K, Borres MP, Högman M, Janson C, et al. Allergen extract vs. component sensitization and airway inflammation, responsiveness and new-onset respiratory disease. *Clin Exp Allergy* 2016;46:730-40.
73. Eder K, Becker S, San Nicoló M, Berghaus A, Gröger M. Usefulness of component resolved analysis of cat allergy in routine clinical practice. *Allergy Asthma Clin Immunol* 2016;12:58.
74. Klemans RJB, Otte D, Knol M, Knol EF, Meijer Y, Gmelig-Meyling FHJ, et al. The diagnostic value of specific IgE to Ara h 2 to predict peanut allergy in children is comparable to a validated and updated diagnostic prediction model. *J Allergy Clin Immunol* 2013;131:157-63.
75. Masthoff L, Mattsson L, Zuidmeer-Jongejan L, Lidholm J, Andersson K, Akkerdaas JH, et al. Sensitization to Cor a 9 and Cor a 14 is highly specific for a hazelnut allergy with objective symptoms in Dutch children and adults. *J Allergy Clin Immunol* 2013;132:393-9.
76. Eller E, Mortz CG, Bindslev-Jensen C. Cor a 14 is the superior serological marker for hazelnut allergy in children, independent of concomitant peanut allergy. *Allergy* 2016;71:556-62.
77. Savvatanos S, Konstantinopoulos AP, Borgà A, Stavroulakis G, Lidholm J, Borres MP, et al. Sensitization to cashew nut 2S albumin, Ana o 3, is highly predictive of cashew and pistachio allergy in Greek children. *J Allergy Clin Immunol* 2015;136:192-4.
78. Lange L, Lasota L, Finger A, Vlajnic D, Büsing S, Meister J, et al. Ana o 3-specific IgE is a good predictor for clinically relevant cashew allergy in children. *Allergy* 2017;72:598-603.
79. Grönlund H, Saarne T, Gafvelin G, van Hage M. The major cat allergen, Fel d 1, in diagnosis and therapy. *Int Arch Allergy Immunol* 2010;151:265-74.
80. Hatzler L, Panetta V, Lau S, Wagner P, Bergmann RL, Illi S, et al. Molecular spreading and predictive value of preclinical IgE response to *Phleum pratense* in children with hay fever. *J Allergy Clin Immunol* 2012;130:894-901.e5.
81. Tsolakis N, Malinovschi A, Nordvall L, Mattsson L, Lidholm J, Pedroletti C, et al. Sensitization to minor cat allergen components is associated with type-2 biomarkers in young asthmatics. *Clin Exp Allergy* 2018;48:1186-94.
82. Schoos A-MM, Chawes BL, Melén E, Bergström A, Kull I, Wickman M, et al. Sensitization trajectories in childhood revealed by using a cluster analysis. *J Allergy Clin Immunol* 2017;140:1693-9.
83. Schoos A-MM, Kattan JD, Gimenez G, Sampson HA. Sensitization phenotypes based on protein groups and associations to allergic diseases in children. *J Allergy Clin Immunol* 2016;137:1277-80.
84. Simpson A, Lazic N, Belgrave DCM, Johnson P, Bishop C, Mills C, et al. Patterns of IgE responses to multiple allergen components and clinical symptoms at age 11 years. *J Allergy Clin Immunol* 2015;136:1224-31.
85. Perzanowski MS, Ronmark E, James HR, Hedman L, Schuyler AJ, Bjerg A, et al. Relevance of specific IgE antibody titer to the prevalence, severity, and persistence of asthma among 19-year-olds in northern Sweden. *J Allergy Clin Immunol* 2016;138:1582-90.
86. Nordlund B, Konradsen JR, Kull I, Borres MP, Önell A, Hedlin G, et al. IgE antibodies to animal-derived lipocalin, kallikrein and secretoglobulin are markers of bronchial inflammation in severe childhood asthma. *Allergy* 2012;67:661-9.
87. Wisniewski JA, Agrawal R, Minniccozi S, Xin W, Patrie J, Heymann PW, et al. Sensitization to food and inhalant allergens in relation to age and wheeze among children with atopic dermatitis. *Clin Exp Allergy* 2013;43:1160-70.
88. Nwaru BI, Suzuki S, Ekerljung L, Sjölander S, Mincheva R, Rönmark EP, et al. Furry animal allergen component sensitization and clinical outcomes in adult asthma and rhinitis. *J Allergy Clin Immunol Pract* 2019;7:1230-8.e4.
89. Fall T, Ekberg S, Lundholm C, Fang F, Almqvist C. Dog characteristics and future risk of asthma in children growing up with dogs. *Sci Rep* 2018;8:16899.
90. Steering Committee Authors, Review Panel Members. A WAO - ARIA - GA2LEN consensus document on molecular-based allergy diagnosis (PAMD@): Update 2020. *World Allergy Organ J* 2020;13:100091.
91. Satyaraj E, Wedner HJ, Bousquet J. Keep the cat, change the care pathway: a transformational approach to managing Fel d 1, the major cat allergen. *Allergy* 2019;74:5-17.
92. Yamamoto-Hanada K, Borres MP, Åberg MK, Yang L, Fukuie T, Narita M, et al. IgE responses to multiple allergen components among school-aged children in a general population birth cohort in Tokyo. *World Allergy Organ J* 2020;13:100105.
93. Sterner T, Uldahl A, Svensson Å, Borres MP, Sjölander S, Tunsäter A, et al. IgE sensitization in a cohort of adolescents in southern Sweden and its relation to allergic symptoms. *Clin Mol Allergy* 2019;17:6.
94. Roberts G, Fontanella S, Selby A, Howard R, Filippi S, Hedlin G, et al. Connectivity patterns between multiple allergen specific IgE antibodies and their association with severe asthma. *J Allergy Clin Immunol* 2020;146:821-30.
95. Hamilton RG, Hemmer W, Nopp A, Kleine-Tebbe J. Advances in IgE testing for diagnosis of allergic disease. *J Allergy Clin Immunol Pract* 2020;8:2495-504.
96. Goodman RE, Chapman MD, Slater JE. The allergen: sources, extracts, and molecules for diagnosis of allergic disease. *J Allergy Clin Immunol Pract* 2020;8:2506-14.
97. Meiser JB, Nelson HS. Comparing conventional and acetone-precipitated dog allergen extract skin testing. *J Allergy Clin Immunol* 2001;107:744-5.
98. Smith DM, Coop CA. Dog allergen immunotherapy: past, present, and future. *Ann Allergy Asthma Immunol* 2016;116:188-93.
99. Uriarte SA, Sastre J. Subcutaneous immunotherapy with high-dose cat and dog extracts: a real-life study. *J Investig Allergol Clin Immunol* 2020;30:169-74.